# STRATEGIES FOR REDUCING THE NUMBER OF FISH USED IN AQUATIC TOXICITY TESTS





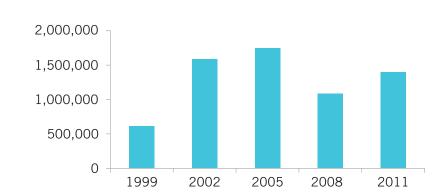
Gilly Stoddart, PhD,<sup>1</sup> Christopher Faßbender, PhD,<sup>1</sup> Patricia Bishop, MSc,<sup>1</sup> Marlies Halder, PhD<sup>2</sup> and Kristie Sullivan, MPH<sup>3</sup>

<sup>1</sup>PETA International Science Consortium Ltd., London, England; PISC@piscltd.org.uk; www.piscltd.org.uk <sup>2</sup>European Commission — Joint Research Centre, Institute for Health and Consumer Protection, STU/EURL ECVAM, Ispra, Italy <sup>3</sup>Physicians Committee for Responsible Medicine, Washington, DC, USA

### INTRODUCTION

In 2011, nearly 180,000 fish were used for toxicological and other safety assessments in Europe. This number is likely to rise in advance of the 2018 REACH deadline.

#### NUMBER OF FISH USED IN THE EUROPEAN UNION FOR SCIENTIFIC PURPOSES2



#### **EXPERIMENTAL DESIGN**

In aquatic toxicity tests, the test chemical is usually added to the tank water. To overcome practical issues associated with testing poorly soluble chemicals, a small volume of solvent is often added. As the solvent can influence the outcome of the study, two controls — one in the presence of and one in the absence of solvent — are currently required, doubling the number of control fish and having significant animal welfare implications.

### NUMBER OF FISH USED IN OECD TEST GUIDELINES FOR REACH

OECD Test Guideline (TG)	# fish per control	# test concentrations	# fish per test concentration replicate	# replicates total	# fish per test if solvent used	# fish saved if no solvent used
TG 203: Fish, Acute Toxicity <sup>3</sup>	7	5	7	1	49	7 (14)
TG 215: Fish, Juvenile Growth <sup>4</sup>	16	5	16	1	112	16 (14)
TG 212: Fish, Short-term Toxicity Test on Embryo and Sac-Fry Stages <sup>5</sup>	30	5	10	3	210	30 (14)
TG 210: Fish, Early-life Stage Toxicity <sup>6</sup>	20	5	20	4	560	80 (14)
TG 305: Bioaccumulation in Fish: Aqueous and Dietary Exposure (e.g. aquatic exposure) <sup>7</sup>	36	2	4	9 time points	114	36 (25)

- Statistical evaluation of historical and simulated data to determine whether one of the controls can be eliminated from aquatic toxicity studies when a solvent is used
- Promote use of advanced techniques to avoid the use of solvents



## **USE ONLY ONE CONTROL WHEN A SOLVENT IS ADDED**

#### **POSSIBLE SOLVENT EFFECTS**

- Subtractive
- Synergistic
- Antagonistic



Current study design (ie, use of two controls) does not allow these interactions between the solvent and test chemical to be determined

If only one control is used when a solvent is necessary, the solvent control is preferred and the water control should be eliminated.

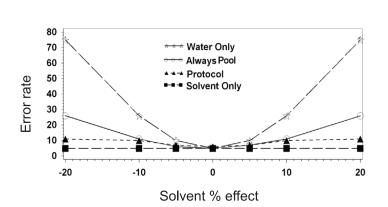
#### CONTROLS USED FOR STATISTICAL ANALYSIS VARY ACCORDING TO REGION,8 GUIDELINE AND TEST RESULTS.

- Water control
- Solvent control<sup>9</sup>
- Always pool water and solvent controls
- Always pool water and solvent controls unless they differ significantly, in which case use only the solvent control, or vice versa.<sup>10</sup>

### STATISTICAL SIMULATIONS<sup>1</sup>

Power to detect a 10% treatment effect <sup>11</sup>									
Solvent	Control used	Power (%)		Solvent	Control	Power (%)			
effect		Two-sided	One-sided	effect	used	Two-sided	One-sided		
	Water	26	38						
0	Solvent	26	38						
	Always pool	35	47						
	Protocol	35	47						
5 ,	Water	51	65	- 5	Water	10	17		
	Solvent	26	38		Solvent	26	38		
	Always pool	49	62		Always pool	21	32		
	Protocol	45	58		Protocol	25	36		
10	Water	75	85	- 10	Water	5	5		
	Solvent	26	38		Solvent	26	38		
	Always pool	62	75		Always pool	11	18		
	Protocol	49	60		Protocol	21	30		

False Positive Error Rates of Control Choices: The probability (%) of indicating a significant treatment effect when there is actually no treatment depends on the choice of control.<sup>11</sup>



## **ALTERNATIVES TO USING SOLVENTS**

#### Direct Dosing<sup>12</sup>

- Prolonged stirring
- High-shear mixing
- Solvent evaporation
- Flow-through systems

- Temperature adjustment
- Large water volumes
- Ultrasonification
- pH adjustment

#### **Passive Dosing**

- Excess chemical added to inert carrier (eg, silica gel) moves passively into water 13,14
- Equilibrium-partitioning occurs between polymer and water
- Losses are compensated for by additional release.

#### Other Generator Systems for Poorly Water-Soluble Substances

- Liquid-liquid saturation units<sup>15</sup>
- Saturator columns (generate saturated solutions without undissolved or emulsified material)

### **SUMMARY**

- The use of solvents for toxicity testing of poorly water-soluble substances in fish currently requires the use of additional fish in solvent controls.
- The need for a water control is scientifically questionable when a solvent is used.

#### Advantages of Using a Single Control

- About 14 to 25% fewer animals when solvent used
- All animals included in analysis
- Fewer resources (eg, time and money)
- Single approach adopted
- Lower false-positive rate

### Disadvantages of Using a Single Control

- There is a decrease in power in some
  - Revisions to TGs and regulatory
  - requirements will be needed.
- A review of historical and simulated data will be needed to determine which controls are required.
- Regulatory harmonisation and mutual acceptance of data will be required to implement changes to controls used in regulatory testing.
- Methods that avoid solvents altogether are available and significantly reduce fish use.
- Animal welfare must be considered when determining control requirements.
- Existing control data are needed from companies to advance this project.

# PETA INTERNATIONAL SCIENCE CONSORTIUM LTD.

### **NEXT STEPS**

#### **FUTURE WORK**

- Collect control data from fish TGs.
- Conduct statistical simulations to determine whether the water control can be eliminated when solvents are used.
- Promote international harmonisation in use of controls.
- Revise and harmonise TGs and regulatory requirements for controls.

#### **CONTRIBUTE DATA TO THIS PROJECT**

- Control data for statistical simulations are needed,
- If you can contribute control data in the presence and absence of a solvent for OECD fish TGs, please go to the QR code or contact GillyS@piscltd.org.uk for more details



### REFERENCES

<sup>1</sup>European Commission. Seventh report on the statistics on the number of animals used for experimental and other scientific purposes in the member states of the European Union. (2013). Available at

http://eur-lex.europa.eu/legal-content/EN/TXT/?uri=CELEX%3A52013DC0859. Accessed 17 May 2016. <sup>2</sup>European Commission. Animals used for scientific purposes: statistical reports. Available at

http://ec.europa.eu/environment/chemicals/lab\_animals/reports\_en.htm. Accessed 17 May 2016.

<sup>3</sup>OECD. (1992). Test Guideline 203: Fish, Acute Toxicity Test.

<sup>4</sup>OECD. (2000). Test Guideline 215: Fish, Juvenile Growth Test.

<sup>5</sup>OECD. (1998). Test Guideline 212: Fish, Short-term Toxicity Test on Embryo and Sac-Fry Stages.

<sup>6</sup>OECD. (2013). *Test Guideline 210: Fish, Early-Life Stage Toxicity Test.* 

<sup>7</sup>OECD. (2012). Test Guideline 305: Bioaccumulation in Fish: Aqueous and Dietary Exposure.

<sup>8</sup>OECD. (2012). Fish Toxicity Testing Framework.

<sup>9</sup>OECD. (2006). Current Approaches in the Statistical Analysis of Ecotoxicity Data: A Guidance to Application. Organisation for Economic Co-operation and Development.

<sup>10</sup>US Environmental Protection Agency. (2008). Guidance for the use of dilution-water (negative) and solvent controls in statistical data analysis for guideline aquatic toxicology studies. Office of Pesticide Programs, Office of Chemical Safety and Pollution Prevention, Washington, DC.

<sup>11</sup>Green JW. (2014). Power and control choice in aquatic experiments with solvents. Ecotoxicol Environ Saf. 102:142-146. <sup>12</sup>Weyman GS, Rufli H, Weltje L, Salinas ER, Hamitou M. (2012). Aquatic toxicity tests with substances that are poorly soluble in water and consequences for environmental risk assessment. Environ Toxicol and Chem, 31(7):1662-1669. <sup>13</sup>Smith KE, Dom N, Blust R, Mayer P. (2010). Controlling and maintaining exposure of hydrophobic organic compounds in aquatic toxicity tests by passive dosing. Aquat Toxicol, 98(1):15-24.

<sup>14</sup>Butler JD, Parkerton TF, Letinski DJ, Bragin GE, Lampi MA, Cooper KR. (2013). A novel passive dosing system for determining the toxicity of phenanthrene to early life stages of zebrafish. Sci Total Environ, 463-464:952-958. <sup>15</sup>Kahl MD, Russom CL, DeFoe DL, Hammermeister DE. (1999). Saturation units for use in aquatic bioassays. Chemosphere,